

Expression of CD123 in B-acute Lymphoblastic leukaemia as a Predictor of Bcr/Abl Rearrangement and Disease Relapse

Bhaumik Prajapati, Birva Raiya, Hemangini Vora*

1

Immunohematology Lab, Cancer Biology Department, The Gujarat Cancer & Research Institute, Ahmedabad, India

* Corresponding Author:
Hemangini Vora,

E-mail:
hemangini.vora@gcriindia.org

ABSTRACT

Introduction: CD123 is the alpha chain of the interleukin 3 receptor (IL-3R) and is generally expressed on hematopoietic progenitor cells, monocytes, B lymphocytes, and endothelial cells. Leukemic stem cells can be detected using CD123, and their usefulness for measuring residual disease and potential involvement in disease relapse is being evaluated. It also regulates the growth, proliferation, survival, and differentiation of hematopoietic cells, along with immunity and inflammatory response.

Materials and Methods: Bone marrow or peripheral blood from 50 B-Acute Lymphoblastic Leukemia (B-ALL) patients were enrolled in the study. CD123 expression was studied by the flow cytometry technique and correlated with clinical and hematological parameters, as well as BCR-ABL status, MRD status, and disease status.

Results: CD123 expression was found positive in 38% of patients. No significant correlation of CD123 expression with clinical and hematological parameters was observed. A significantly higher incidence of CD123 expression was noted in patients with BCR-ABL fusion (70%), relapse patients (67%), and MRD patients (67%).

Conclusion: CD123 can be used to predict BCR-ABL status in B-ALL patients, and it has a potential role in recognizing high-risk relapse and helps to scrutinize high-risk B-ALL patients who benefited from aggressive chemotherapy. Further, higher expression of CD123 in MRD patients can be used to evaluate minimal residual disease in follow-up B-ALL patients.

Keywords: B-Acute Lymphoblastic Leukemia; Flow cytometry; CD123



INTRODUCTION:

Acute lymphoblastic leukemia (ALL) is a malignant disease characterized by the clonal growth of leukemic cells in the bone marrow (BM), lymph nodes, thymus, and spleen. This disease is diverse and classified into multiple subtypes. Acute lymphoblastic leukemia (ALL) accounts for 75% of all leukemias in children under the age of 15, affecting individuals of all ages; however, it is most prevalent among young people. In adults, it is more common in those over 45. B-acute lymphoblastic leukemia/lymphoma (B-ALL) is a hematologic malignancy originating from B-cell progenitors.

CD123 is the alpha chain of the interleukin 3 receptor (IL-3R) and is generally expressed on hematopoietic progenitor cells, monocytes, B lymphocytes, and endothelial cells. Several hematologic neoplasms, including B-ALL, express CD123, but normal hematopoietic stem cells either express it less or don't express it at all [3]. Importantly, it has been reported that leukemic stem cells and more differentiated leukemic blast cells both express CD123 positively [4, 5]. Leukemic stem cells can be detected using CD123, and its usefulness for measuring residual disease and potential involvement in disease relapse is being evaluated [6]. It also regulates the growth, proliferation, survival, and differentiation of hematopoietic cells, along with immunity and inflammatory response [7,8]. This study aimed to assess the pattern of CD123 expression in B-ALL patients.

Materials and Methods:

Patient Characteristics

In this prospective study, 50 B-cell Acute Lymphoblastic Leukemia (B-ALL) patient samples were collected at The Gujarat Cancer & Research Institute (G.C.R.I.) from January 2023 to April 2023. First differential counts at diagnosis and clinicopathological data such as age, gender, karyotype, and bone marrow were recorded from available hospital records files maintained at the Institutional Medical Record Department. Patients provided general consent to use their sample for the study. This study was approved by the Institutional Scientific Review Board and Ethics Committee.

Sample collection

Bone marrow or peripheral blood samples of 50 patients (25 newly diagnosed and 25 follow-up patients) were collected in Ethylenediamine Tetra Acetic Acid (EDTA) vacuette.

Sample preparation

100 µl BM/PB samples were stained with 5 µl CD45 (V500c) + 10 µl CD123 (FITC) monoclonal antibodies, lysed with 2 ml RBC lysing solution (1:10 Dilution), washed with 2 ml Phosphate buffer saline (PBS), and then resuspended with 500 µl PBS. Then, subjected to a Flow Cytometer within 72 hours.

Sample acquisition

Sample acquisition is done in the FACSCanto II (Flow Cytometer) instrument using FACSDiva software. During acquisition, the global worksheet is kept open and a total of 1,00,000 cells or events are acquired.

Data analysis

For analysis, the global worksheet is changed to a normal worksheet. CD123-positive cells are selected from the Dim CD45 population. The percentage of each positive population is noted from the "population hierarchy" table. A histogram is plotted and gated to find the CD123 MFI (median fluorescence intensity) value of leukemic blasts.

Statistical analysis

Statistical analysis was carried out using SPSS statistical software version 27. A receiver operating characteristic (ROC) curve was generated to determine the sensitivity and specificity of the marker. Pearson's chi-square test with Pearson's correlation coefficient (r) was used to assess correlation and significance between two parameters. P values ≤ 0.05 were considered to be statistically significant.

RESULTS:

Out of 50 B-ALL patients, CD123 expression was found positive in 19(38%) patients and negative in 31 (62%) patients (Figures 1 & 2). The expression was further correlated with clinical and hematological parameters. In correlation with clinical parameters, a trend of higher incidence of CD123 expression was found in male, absence of hepatomegaly and lymphadenopathy as compared to their counterparts. In correlation with hematological parameters, a trend of higher incidence of CD123 expression was found in B-ALL patients with higher blast cell count, lymphocytes, and platelets as compared to their counterparts. No significant correlation of CD123 expression with other clinical and hematological parameters was observed (Table 1).

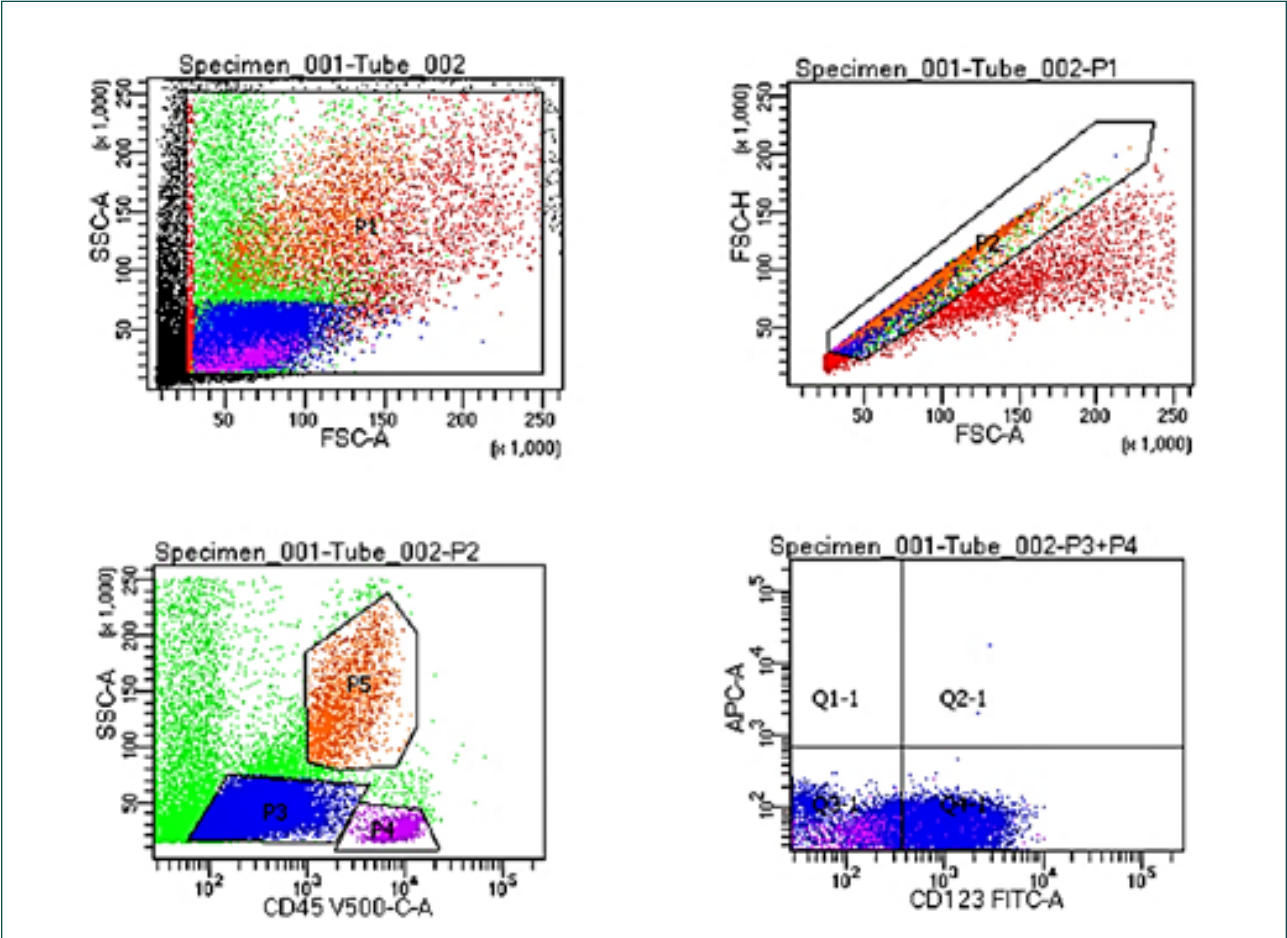


Figure 1. Expression of CD123 in B-ALL

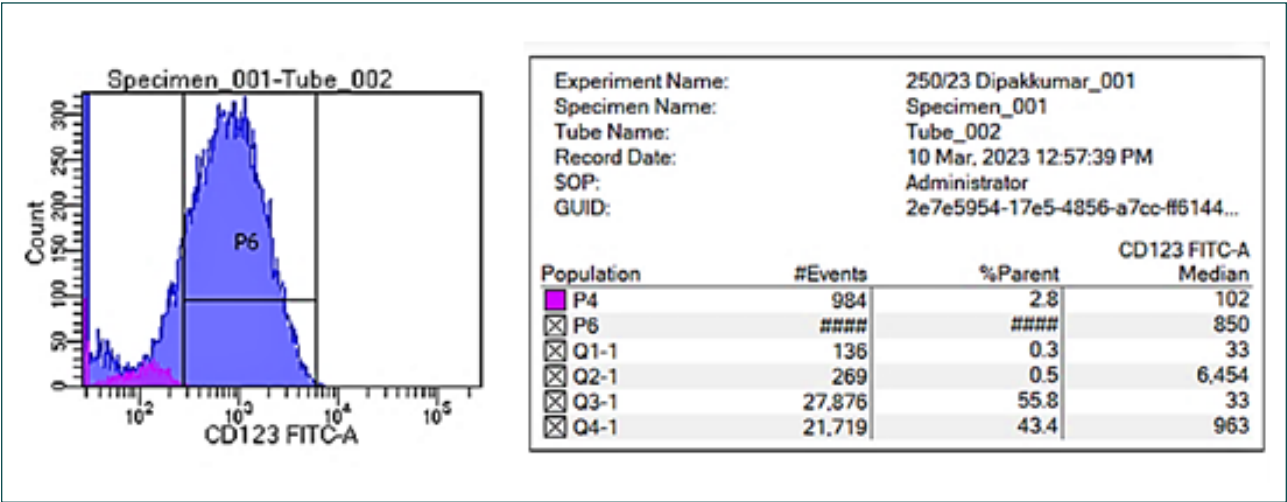


Figure 2. Median fluorescence intensity of CD123 in B-lymphoblasts

Table 1. Correlation of CD123 expression with clinical and hematological parameters

Parameters	Total Patients N (%)	CD123 Negative N (%)	CD123 Positive N (%)	χ^2	R	P
Age	50(100)	31(62)	19(38)	0.08	0.04	0.77
Adult	25(50)	15(60)	10(40)			
Pediatric	25(50)	16(64)	9(36)			
Gender	50(100)	31(62)	19(38)	1.24	0.16	0.26
Male	32(64)	18(56)	14(44)			
Female	18(36)	13(72)	5(28)			
Splenomegaly	40(100)	24(60)	16(40)	0.07	-0.04	0.79
Positive	21(52)	13(62)	8(38)			
Negative	19(48)	11(58)	8(42)			
Hepatomegaly	40(100)	24(60)	16(40)	2.06	-0.23	0.15
Positive	23(58)	16(70)	7(30)			
Negative	17(42)	8(47)	9(53)			
Lymphadenopathy	21(100)	10(48)	11(52)	0.40	-0.14	0.52
Positive	5(24)	3(60)	2(40)			
Negative	16(76)	7(44)	9(56)			
Hemoglobin (gm/dl)	50(100)	31(62)	19(38)	0.19	-0.06	0.66
>8.9	23(46)	15(65)	8(35)			
≤8.9	27(54)	16(59)	11(41)			
RBC (cells/ μ l)	50(100)	31(62)	19(38)	0.26	0.07	0.61
>3.27*10 ⁶	24(48)	14(58)	10(42)			
≤3.27*10 ⁶	26(52)	17(65)	9(35)			
WBC (cells/ μ l)	50(100)	31(62)	19(38)	0.08	-0.04	0.77
>7.295*10 ³	25(50)	16(64)	9(36)			
≤7.295*10 ³	25(50)	15(60)	10(40)			
Platelets (cells/ μ l)	50(100)	31(62)	19(38)	1.20	0.15	0.27
>34x10 ³	24(48)	13(54)	11(46)			
≤34x10 ³	26(52)	18(69)	8(31)			
Lymphocytes (%)	50(100)	31(62)	19(38)	3.31	-0.26	0.07
>30	24(48)	18(75)	6(25)			
≤30	26(52)	13(50)	13(50)			
Polymorphs (%)	49(100)	30(61)	19(39)	0.002	0.007	0.96
>25	23(47)	14(61)	9(39)			
≤25	26(53)	16(61)	10(39)			
Blast cells (%)	31(100)	22(71)	9(29)	1.70	0.23	0.19
>52	15(48)	9(60)	6(40)			
≤52	16(52)	13(81)	3(19)			

(χ^2 = Chi-square; R = Pearson's correlation coefficient; P = p-value)

CD123 expression concerning BCR-ABL status

Correlation of CD123 expression was evaluated with BCR-ABL status, as shown in Table 2. BCR-ABL fusion was detected as positive in 10 out of 45 B-ALL patients. A higher incidence of CD123-positive expression (7/10, 70%) and low expression of CD123-negative expression (3/10, 30%) was seen in the BCR-ABL positive group ($P = 0.01$). A lower incidence of CD123 positive expression (29%, 10/35) and a higher incidence of CD123 negative expression (25/35, 71%) was seen in BCR-ABL negative group ($P = 0.01$) (Table.2). ROC curve analysis (AUC 0.723) suggests patients with CD123 MFI more than 130 have higher chance of having BCR-ABL fusion (Figure.3).

CD123 expression concerning MRD status

Correlation of CD123 expression was evaluated with MRD status as shown in Table 2. The MRD Status of 20 patients was evaluated, and it was detected as positive in 9 out of 20 B-ALL patients, and a higher incidence of CD123-positive expression was seen in the MRD-positive group (67%, 6/9) ($P = 0.007$) (Figure 4). A lower

incidence of CD123-positive expression was seen in the MRD negative group (9%, 1/11) ($P = 0.007$) (Table 2).

CD123 expression concerning Disease status

Disease status of 30 patients was available, out of which 20% (6/30) showed disease relapse and 80% (24/30) showed disease remission. CD123 expression was found positive in 33% (10/30), with 4 relapse patients and 6 remission patients. ($P = 0.05$). CD123 expression was found negative in 67% (20) with 2 relapse patients and 18 remission patients ($P = 0.05$) (Table 2). ROC curve analysis (AUC 0.820) suggests patients with CD123 MFI more than 108 have a higher chance of having disease relapse (Figure 5)

DISCUSSION:

A clonal hematologic malignancy - acute lymphoblastic leukaemia (ALL) develops from T- or B-lymphoid progenitor cells. In the present study, CD123 expression was found in 38% (19/50) of the studied B-ALL patients. This incidence is lower than that reported by Angelova E et al [3] (164/183, 89.6%); Djokic M et al [9] (98/119,

Table 2. Correlation of CD123 expression with BCR-ABL status

BCR-ABL Status N (%)	CD123 Expression (N%)		χ^2	R	P
	Positive N (%)	Negative N (%)			
Positive 10(22)	7(70)	3(30)	5.68	0.35	0.01
Negative 35(78)	10(29)	25(71)			
Total 45(100)	17(38)	28(62)			
MRD Status N (%)			7.21	0.60	0.007
Positive 9(45)	6(67)	3(33)			
Negative 11(55)	1(9)	10(91)			
Total 20(100)	7(35)	13(65)			
Disease Status N (%)			3.75	0.35	0.05
Relapse	4(67)	2(33)			
Remission	6(25)	18(75)			
Total	10(33)	20(67)			

(χ^2 = Chi-square; R = Pearson's correlation coefficient; P = p-value)

82%), Hassanein NM et al [10] (45/50, 89%) and Bras AE et al [11] who detected CD123 expression in 85% of B-cell precursor (BCP) ALL cases (224/ 262). In the present study, out of 50 B-ALL patients, CD123 expression was found in 44% of male (14/32) and 28% of female (5/18) patients. The study by Li Z et al [12] has shown that out of 328 pediatric B-ALL patients, CD123 expression was found in 55% (105/190) male and 51% female (70/138) patients. In the current study, expression of CD123 was not significantly correlated with the haematological parameters such as haemoglobin, RBC, WBC, platelets, lymphocytes, polymorphs, and blast cells. Similarly, in the study by Aref et al [13], no significant correlation of CD123 expression was observed with haemoglobin, WBC, platelets, and blast cells count.

3-5% of children and 20-30% of adults have Philadelphia-positive (Ph+) B-ALL, and the frequency rises to roughly 50% in people over the age of 50. In the present study, the correlation of CD123 expression was evaluated with BCR-ABL fusion status. There was a strong association between CD123-positive expression and BCR-ABL BCR-ABL-positive group ($p = 0.017$). BCR-ABL fusion was detected in 22% (10/45) of B-ALL cases, with 6% in children and 16% in adults. This finding is lower than that reported by Gadhia P et al [14] (33.3%); Aref S et al [13] (30%), and higher than that reported by Owaidah TM et al [15] (17.5% (18/103)).

In the present study, follow-up samples of 20 B-ALL patients were evaluated, and 9 patients showed minimal residual disease (MRD). Out of these 9 patients, 6 (67%) showed CD123 expression, this incidence is nearly

similar to the study by Li Z et al. [12] (61.3%). Das et al [6] reported that the presence of CD123 expression at baseline was substantially more frequently related to MRD-positive status ($P < 0.001$ for 10% or 20% and $P = 0.005$ for 5% of blasts expressing CD123).

In the present study, the disease status of 30 B-ALL patients was evaluated, and out of these, 6 patients showed relapse and 24 patients showed remission. Out of 6 relapse patients, CD123 expression was seen in 4 (66.6%) relapse patients, and out of 24 remission patients, CD123 expression was seen in 6 (25%) remission patients. In contrast to this finding, Li Z et al [12] reported that expression of CD123 was seen in 39.3% relapse patients and 53.3% remission patients. This difference may be due to the enrolled only pediatric B-ALL patients in their study. Thus, this study demonstrated that high expression of CD123 was correlated with BCR-ABL positivity, MRD positivity, and disease relapse.

CONCLUSION:

CD123 can be used to predict BCR-ABL status in B-ALL patients, and it has the potential role to recognize high risk of relapse and helps to scrutinize high-risk B-ALL patients who benefited from aggressive chemotherapy. Further, higher expression of CD123 in MRD patients can be used to evaluate minimal residual disease in follow-up B-ALL patients.

CONFLICT OF INTEREST:

The authors declare no conflicts of interest.

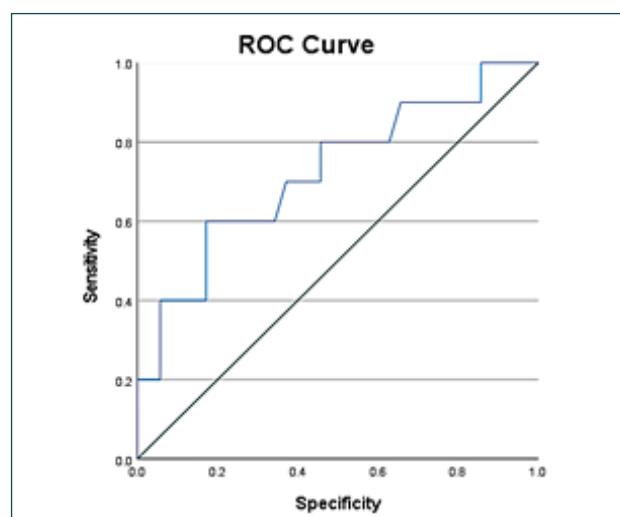


Figure 3. ROC Curve showing CD123 expression with BCR-ABL

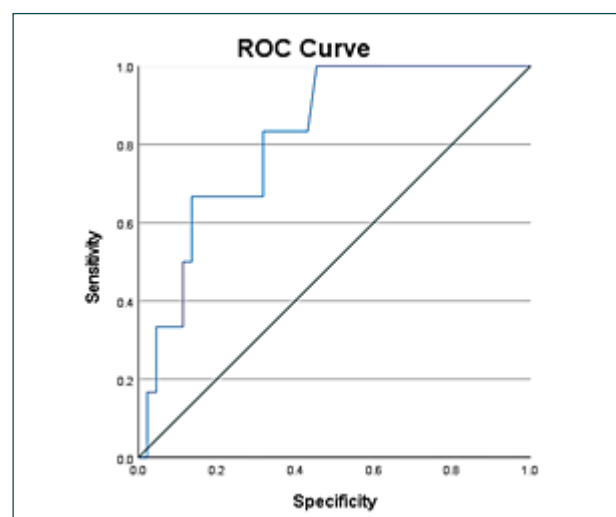


Figure 5. ROC curve showing CD123 expression with disease status

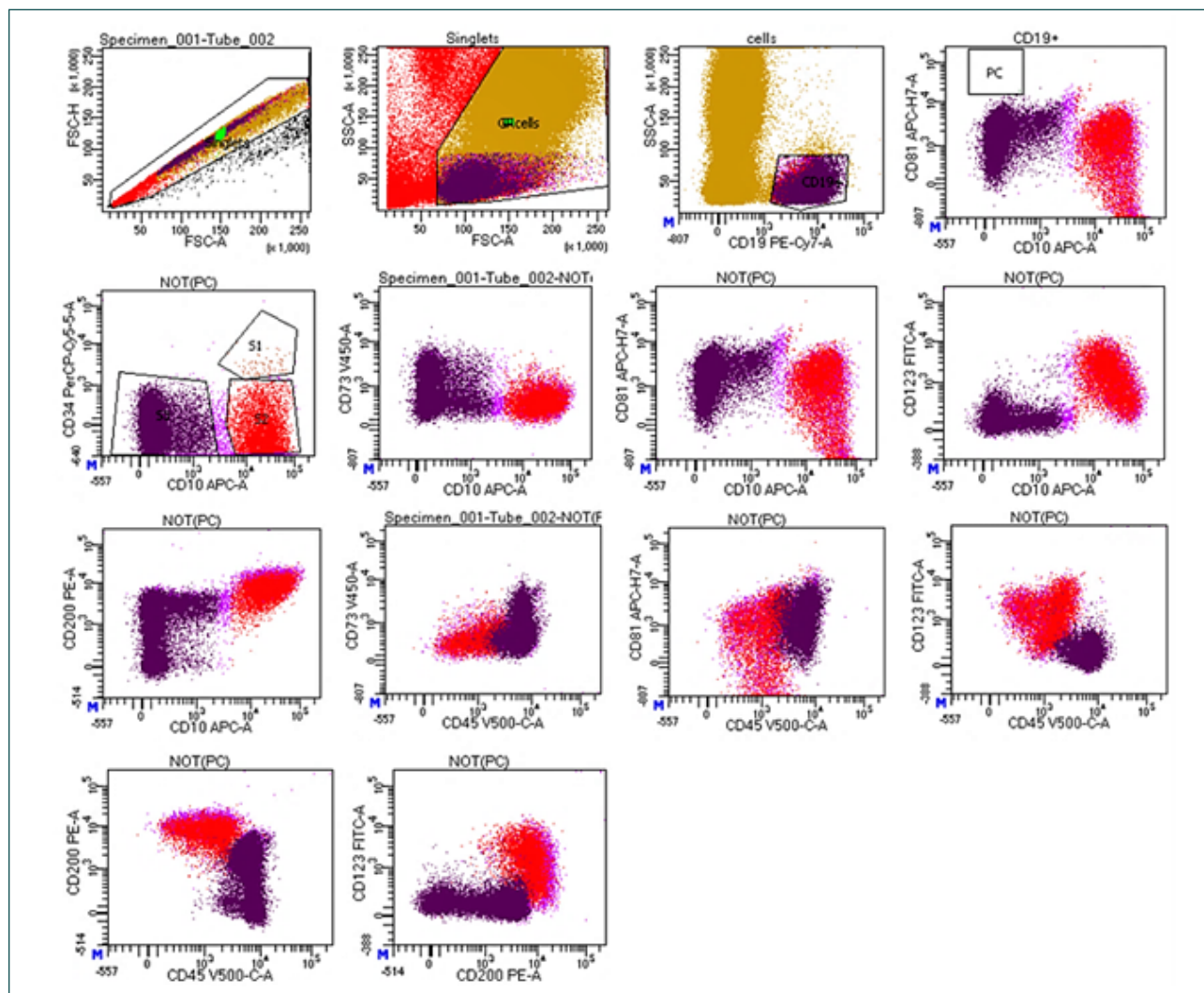


Figure 4. Aberrant CD123 expression in B-ALL is useful in MRD detection along with other markers

References:

1. Faderl S, Jeha S, Kantarjian HM. The biology and therapy of adult acute lymphoblastic leukemia. *Cancer: Interdisciplinary International Journal of the American Cancer Society*. 2003 Oct 1;98(7):1337-54.
2. Campo E, Harris NL, Jaffe ES, Pileri SA, Stein H, Thiele J, Vardiman JW. WHO classification of tumours of haematopoietic and lymphoid tissues. Swerdlow SH, editor. Lyon: International agency for research on cancer; 2008 Sep 20.
3. Angelova E, Audette C, Kovtun Y, Daver N, Wang SA, Pierce S, Konoplev SN, Khogeer H, Jorgensen JL, Konopleva M, Zweidler-McKay PA. CD123 expression patterns and selective targeting with a CD123-targeted antibody-drug conjugate (IMGN632) in acute lymphoblastic leukemia. *Haematologica*. 2019 Apr;104(4):749.
4. U. Testa, E. Pelosi, and G. Castelli, "CD123 as a therapeutic target in the treatment of hematological malignancies," *Cancers*, vol. 11, no. 9, p. 1358, 2019.
5. Ruella M, Barrett DM, Kenderian SS, Shestova O, Hofmann TJ, Perazzelli J, Klichinsky M, Aikawa V, Nazimuddin F, Kozlowski M, Scholler J. Dual CD19 and CD123 targeting prevents antigen-loss relapses

- after CD19-directed immunotherapies. *The Journal of clinical investigation*. 2016 Oct 3;126(10):3814-26.
6. Das N, Gupta R, Gupta SK, Bakhshi S, Malhotra A, Rai S, Singh S, Prajapati VK, Sahoo RK, Gogia A, Sharma A. A real-world perspective of CD123 expression in acute leukemia as promising biomarker to predict treatment outcome in B-ALL and AML. *Clinical Lymphoma Myeloma and Leukemia*. 2020 Oct 1;20(10):e673-84.
 7. Broughton SE, Dhagat U, Hercus TR, Nero TL, Grimbaldston MA, Bonder CS, Lopez AF, Parker MW. The GM-CSF/IL-3/IL-5 cytokine receptor family: from ligand recognition to initiation of signaling. *Immunological reviews*. 2012 Nov;250(1):277-302.
 8. Guthridge MA, Stomski FC, Thomas D, Woodcock JM, Bagley CJ, Berndt MC, Lopez AF. Mechanism of activation of the GM-CSF, IL-3, and IL-5 family of receptors. *Stem cells*. 1998 Sep;16(5):301-13.
 9. Djokic M, Björklund E, Blennow E, Mazur J, Söderhäll S, Porwit A. Overexpression of CD123 correlates with the hyperdiploid genotype in acute lymphoblastic leukemia. *haematologica*. 2009 Jul;94(7):1016.
 10. Hassanein NM, Alcancia F, Perkinson KR, Buckley PJ, Lagoo AS. Distinct expression patterns of CD123 and CD34 on normal bone marrow B-cell precursors ("hematogones") and B lymphoblastic leukemia blasts. *American journal of clinical pathology*. 2009 Oct 1;132(4):573-80.
 11. Bras AE, de Haas V, van Stigt A, Jongen-Lavrencic M, Beverloo HB, Te Marvelde JG, Zwaan CM, van Dongen JJ, Leusen JH, van der Velden VH. CD123 expression levels in 846 acute leukemia patients based on standardized immunophenotyping. *Cytometry Part B: Clinical Cytometry*. 2019 Mar;96(2):134-42.
 12. Li Z, Chu X, Gao L, Ling J, Xiao P, Lu J, Wang Y, He H, Li J, Hu Y, Li J. High expression of interleukin-3 receptor alpha chain (CD123) predicts favorable outcome in pediatric B-cell acute lymphoblastic leukemia lacking prognosis-defining genomic aberrations. *Frontiers in Oncology*. 2021 Mar 16;11:614420.
 13. Aref S, El Agdar M, Khaled N, Ibrahim L, El-Ghohemy MS. Clinical Impact of CD25/CD123 Coexpression in Adult B-Cell Acute Lymphoblastic Leukemia Patients. *Advances in Hematology*. 2020 May 20;2020.
 14. Gadhia P, Parek N, Chavda P, Bhatia G, Vaniawala S. Cytogenetic findings of patients with acute lymphoblastic leukemia in west Indian region. *International Journal of Advanced Research*. 2018;6(2):2320-407.
 15. Owaidah TM, Rawas FI, Elcum NB. Expression of CD66c and CD25 in acute lymphoblastic leukemia as a predictor of the presence of BCR/ABL rearrangement. *Hematology/Oncology and Stem Cell Therapy*. 2008 Jan 1;1(1):34-7